

What is claimed is:

1. A method of producing aflibercept, comprising:
 - (a) producing a clarified harvest of cells cultured in a chemically defined medium (CDM);
 - (b) binding aflibercept from said clarified harvest to a Protein A resin;
 - (c) eluting said aflibercept of step (b) forming an affinity eluate, wherein said eluate has a first color;
 - (d) subjecting said eluate comprising aflibercept to anion exchange chromatography (AEX); and
 - (e) collecting a flowthrough fraction, wherein said flowthrough fraction has a second color, and wherein said first color of said affinity eluate is a more intense yellow brown color than said second color of said flowthrough fraction when said eluate and flowthrough protein concentrations are normalized.
2. The method of claim 1, wherein said first color has a b* value ranging from about 2.0 to about 20.0 when said protein concentration is normalized to 10.0 g/L.
3. The method of claim 1, wherein said second color has a b* value ranging from about 0.5 to about 5.0 when said protein concentration is normalized to 10.0 g/L.
4. The method of claim 1, wherein said cell is selected from a group consisting of CHO, NS0, Sp2/0, embryonic kidney cells and BHK.
5. The method of claim 1, wherein said clarified harvest comprises one or more aflibercept variants, wherein said variants have at least one oxidized amino acid residue.
6. The method of claim 5, wherein said oxidized amino acid residue is selected from group consisting of methionine, tryptophan, histidine, phenylalanine, tyrosine and a combination thereof.
7. The method of claim 6, wherein said oxidized amino acid residue is histidine.
8. The method of claim 6, wherein said oxidized amino acid residue is tryptophan.
9. The method of claim 1, wherein said AEX column comprises an anionic exchange substituent including diethylaminoethyl (DEAE), quaternary aminoethyl (QAE) and quaternary amine (Q) groups.
10. The method of claim 5, wherein said aflibercept variant is selected from an amino acid residue on a polypeptide having an amino acid sequence as set forth in the group consisting of: SEQ ID NO.: 17, SEQ ID NO.: 18, SEQ ID NO.: 19, SEQ ID NO.: 20, SEQ ID NO.: 21, SEQ ID NO.: 22, SEQ ID NO.: 23, SEQ ID NO.: 56, SEQ ID NO.: 62, SEQ ID NO.: 63, SEQ ID NO.: 64, SEQ ID NO.: 65, SEQ ID NO.: 66, SEQ ID NO.: 67, SEQ ID NO.: 68, SEQ ID NO.: 69, SEQ ID NO.: 70, SEQ ID NO.: 71 and combinations thereof.
11. The method of claim 1, further comprising after binding aflibercept from said clarified harvest, subjecting said aflibercept to one or more further chromatographic steps selected from the group consisting of: cation exchange chromatography, hydrophobic interactive chromatography, size exclusion chromatography and a combination thereof.
12. A method of producing aflibercept from a clarified harvest of a cell cultured in a chemically defined medium (CDM), comprising:
 - (a) binding aflibercept from said clarified harvest to a Protein A resin;
 - (b) eluting said aflibercept of step (a) forming an affinity eluate, wherein said eluate comprises acidic species of aflibercept;
 - (c) subjecting said eluted aflibercept to anion exchange (AEX) chromatography; and
 - (d) collecting one or more flowthrough fractions, and wherein the percent of acidic species of aflibercept in said affinity eluate is greater than the percent of acidic species of aflibercept in said one or more flowthrough fractions when the concentrations of protein in said eluate and flowthrough fractions are normalized, and wherein said acidic species of aflibercept correspond to peaks that elute earlier than a main peak in a cation exchange chromatography (CEX) chromatogram of aflibercept, and wherein the chromatogram is generated using a first mobile phase of 20 mM 2-(N-morpholino) ethanesulfonic acid (MES), pH 5.7 and a second mobile phase of 40 mM sodium phosphate, 100 mM sodium chloride pH 9.0 (Mobile phase B), and wherein the chromatogram is generated using detection at 280 nm.
13. The method of claim 12, wherein said acidic species of aflibercept in said affinity eluate are reduced by at least ten percent compared to said flowthrough fraction when the concentrations of said affinity eluate and flowthrough protein are normalized.
14. The method of claim 12, wherein said aflibercept from the one or more flowthrough fractions comprises less than 20% total acidic species of aflibercept.
15. The method of claim 12, wherein said acidic species of aflibercept comprises aflibercept having at least one oxidized amino acid residue selected from group consisting of methionine, tryptophan, histidine, phenylalanine, tyrosine and a combination thereof.
16. The method of claim 12, wherein the pH of both the equilibration and wash buffers for the AEX column are from about 8.30 to about 8.60.
17. The method of claim 12, wherein the conductivity of both the equilibration and wash buffers for said AEX column can be from about 1.50 to about 3.0 mS/cm.
18. The method of claim 12, further comprising after binding aflibercept from said clarified harvest, subjecting aflibercept to one or more further chromatographic steps selected from the group consisting of: cation exchange chromatography (CEX), hydrophobic interactive chromatography, size exclusion chromatography and a combination thereof.
19. A method of producing aflibercept from a clarified harvest of a cell cultured in a chemically defined medium (CDM), comprising:
 - (a) binding aflibercept from said clarified harvest to a Protein A resin;
 - (b) eluting said aflibercept of step (a) forming an affinity eluate, wherein said eluate comprises oxidized species of aflibercept;
 - (c) subjecting said eluted aflibercept to anion exchange (AEX) chromatography; and
 - (d) collecting a flowthrough fraction, wherein the percent of oxidized species of aflibercept in said affinity eluate is greater than the percent of oxidized species of aflibercept in said flowthrough fraction when the concentrations of protein in said eluate and flowthrough fraction are normalized, and wherein said oxidized species of aflibercept is measured by subjecting said affinity eluate and said flowthrough fractions to digestion, followed by their analysis using reverse-phase ultra-performance chromatography (UPLC), detection at wavelengths of 280 nm, 320 nm and 350 nm and